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# Physicochemical properties of different organic substrate mixtures and their effects on growth and yield of sweet pepper (*Capsicum annuum* L.) cultivars

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## Abstract

Carbonized rice husk, coco peat and sawdust are considered as good growing substrate components, but high water holding capacity causes poor air-water relationship, thus affecting oxygen diffusion to roots. Incorporation of coarser materials into these substrate components could improve aeration status. The present experiment aimed to assess the physicochemical properties of four growing substrate mixtures ( $M_1 = 50\%$  coco peat + 45% bora + 5% shodo,  $M_2 = 50\%$  coco peat + 45% perlite + 5% shodo,  $M_3 = 50\%$  carbonized rice husk + 45% perlite + 5% shodo and  $M_4 = 50\%$  sawdust + 45% perlite + 5% shodo) and their effects on growth and yield of two sweet pepper cultivars ( $V_1 =$  'Papari new-E-red',  $V_2 =$  'AVRDC PP045-6006'). Results revealed that pH, electrical conductivity (EC), air-filled

porosity (AFP) and wettability were higher in  $M_3$ , whereas bulk density and dry weight were higher in  $M_1$  and the lowest in  $M_3$ . Improved properties of  $M_3$  and  $M_2$  positively reflected in growth, dry weight, yield and mineral compositions in leaves of sweet pepper. Furthermore,  $V_2$  showed significantly better growth and higher yield as compared to  $V_1$  cultivar. Therefore, we concluded that incorporation of coarser materials improved physicochemical properties of carbonized rice husk ( $M_3$ ) followed by coco peat ( $M_2$ ) that positively influenced the growth and yield of sweet pepper cv. 'Papari new-E-red'.

**Keywords:** Carbonized rice husk; Coco peat; Sawdust, Growth; Dry weight, Tissue mineral contents

## 1. Introduction

Use of suitable growing substrate is essential for production of high quality horticultural crops. It directly affects the development and later maintenance of the functional rooting system. A good growing medium provides a sufficient support to the plant, serves as reservoir for nutrients and water, allows oxygen diffusion to the roots and permits gaseous exchange between the roots and atmosphere outside the root substrate (Abad et al., 2002; Argo, 1998; Bunt, 1988; Richards and Beardsell, 1986). Many soilless materials are widely available in the tropics, for example coco peat, carbonized rice husk, and sawdust, etc. These materials are mainly agricultural byproducts obtained after the extraction of fiber from the coconut husk, paddy and saw mills and maybe used as horticultural growing substrates. As a growing medium, coco peat can be used to produce a number of crop species with acceptable quality in the tropics (Yahya et al., 1997). Coco peat is considered as a good growing substrate (Abad et al., 2002). However, coco peat has been recognized to have high water holding capacity which causes poor air-water relationship, leading to low aeration within the medium and it affects the oxygen diffusion to the roots.

Carbonized rice husk maybe used as horticultural growing substrate although it has the problem of air-water relationship. Rice hull is often incinerated to form fine charcoal-like dust. When it is used as a component for growing substrate, it might behave like fine sand. However, it is lighter and sterile and may contain some nutritional elements. Carbonized rice husk induced faster cell division and differentiation for root formation (Moe, 1988) and it was

the best growing substrate for chrysanthemum cutting (Budiartoa et al., 2006). Aside its use in nursery production, it maybe used as soilless growing media for sweet pepper production.

Sawdust is used as growing substrate and is available in almost all over the world and it could be renewable. Wood residues (i.e., sawdust and bark) have been used in containers for growing ornamentals (Klett et al., 1972). But microorganisms involved in decomposition of raw wood residues are more efficient than higher plants in nitrogen absorption and assimilation (Alexander, 1961). Large amount of nitrogen must, therefore, be added to wood residues used as media to grow plants. This problem can be solved, however, by composting residues before using them for growing substrates (Still et al., 1974). Furthermore, decomposition of old sawdust is not necessary for using as growing substrates. However, physical and chemical properties of these substrate components are highly dependent on their processing technique and handling. It is desirable to improve physical and chemical properties of them before using as growing substrates.

Incorporation of coarser materials into the substrate components could improve the aeration and drainage status of the substrate mixtures (Bunt, 1988; Richards and Beardsell, 1986; Sambo et al., 2008). Perlite (Islam, 2008; Sambo et al., 2008), bora (volcanic soil), and shodo (burnt clay loam soil), the possible coarser materials can be used to improve the air-water relationship of the substrate components. Possible combinations of different components can improve the physical and chemical properties of growing substrate. Furthermore, a suitable combination of different growing substrate components positively influences the growth and yield of horticultural crop production like sweet pepper as a test crop. Thus, the objectives of the study were to assess the chemical and physical properties of different substrate mixtures, and their effects on growth, plant dry weight, yield, and mineral composition in leaves of sweet pepper.

## 2. Materials and methods

#### Plant materials and growing condition

The experiment was conducted at the greenhouse of the Experimental Farm, University of Miyazaki, Japan from November 2010 to June 2011. The transplanting and final harvesting dates were 25th November 2010 and 10th June 2011, respectively. Seeds of sweet pepper cv. 'Papri new-E-red' (Marutane Seed Co., Kyoto, Japan) and 'AVRDC PP046-6006' (AVRDC-The World Vegetable Centre, Taiwan) were selected on the basis of yield

performances under high temperature condition (Rahman and Inden, 2012). Two 18 cm, sixth leaf stage 8-week-old seedlings were transplanted 20 cm apart into 40 L containers containing different mixtures Each row consisted of 18 containers and treated as a replication. Two edge rows were grown to reduce the border effects. The first 3-flowers were removed from all plants at anthesis to promote vegetative growth. Plants were pruned to form four main stems. The nutrient solution was applied to the crop by ultra drip irrigation tube for each replication and treatment. The nutrient solution applied was measured with a flow meter, installed in a water delivery unit designed for independent control of each treatment. A standard nutrient solution was used in this experiment according to Rahman and Inden (2012). The pH and electrical conductivity (EC) of 6.0 and 2.8 dS·m<sup>-1</sup>, respectively were maintained during experiment. The nutrient composition of the solution was NO<sub>3</sub>-N, P, K, Ca, Mg, and S of 17.0, 7.8, 8.7, 9.9, 6.0 and 6.0 me L<sup>-1</sup>, respectively, and Fe, B, Zn, Cu, Mo, and Mn of 3.0, 0.5, 0.1, 0.03, 0.025 and 1.0 mg L<sup>-1</sup>, respectively. The average minimum and maximum temperatures during the cultivation period were  $18\pm 2$  °C and  $22\pm 2$  °C, respectively.

#### Experimental design and treatments

The experimental design was a  $4\times2$  factorial design with three replications. Two factors of this experiment were four substrate mixtures (viz.,  $M_1 = 50\%$  coco peat + 45% bora + 5% shodo,  $M_2=50\%$  coco peat + 45% perlite + 5% shodo,  $M_3=50\%$  carbonized rice husk + 45% perlite + 5% shodo, and  $M_4 = 50\%$  sawdust + 45% perlite + 5% shodo), and two sweet pepper cultivars (viz.,  $V_1$  = 'Papri new-E-red' and  $V_2$  = 'AVRDC PP046-6006'). Four substrate mixtures ( $M_1$ ,  $M_2$ ,  $M_3$  and  $M_4$ ) were used as completely randomized design with four replications for determination of physical and chemical properties.

#### Data collection

#### **Properties of growing substrate**

The selected properties of growing substrate, namely initial pH and EC, bulk density, water retention, wettability, air-filled porosity and dry weight of substrate mixture were measured. Samples of substrate mixtures were collected accordingly after preparation and then put to specific determination of the properties.

#### pH and EC

The pH and EC values for all media before planting were determined according to Yahya et al.

(2009). For pH, 10 g of media was mixed with 50 mL distilled water, agitated for 30 min and left standing for 24 h. For EC, 40 g of media was mixed with 80 mL distilled water, shaken for 15 min and left for 60 min. The mixtures were filtered before the measurements. A pH meter (pH/COND Meter, D-54, Horiba Ltd., Kyoto, Japan) was used to measure pH and EC.

## Bulk density $(g \cdot cm^{-3})$

Bulk density was determined by using the core method (Teh and Jamal, 2006). In brief, the substrate mixtures in the core rings were saturated by allowing water to diffuse into the substrate for two days. The samples were oven-dried at 105°C for 24 h and recorded their weights. The bulk density ( $\rho$ b) was calculated as the following formula,  $\rho b = (W_b-W_r)/(\pi h_t d^2/4)$ , where  $W_b$  is the weight of oven dried substrate mixture and core ring (g),  $W_r$  is the weight of the core ring (g),  $h_t$  is the core ring height (cm) and d is the core ring diameter (cm).

### Water retention (%)

Water retention was measured by using 1 L plastic pot with hole at the bottom. A blotting paper was placed to cover the hole at the bottom and the empty pot with the blotting paper was weighed. After filling the pot with the oven dried substrate mixtures it was reweighed with substrate mixtures. The pots were put in the plastic trays with water until the mixtures were wet through 24 h. Then the pots were stood in a beaker so that the water could be drained out freely by gravity. Finally the pots were weighted and the water retention was measured using the following formula, water retention (%) =  $\{(Ws-W_d)/W_d\} \times 100$ , where  $W_s$ -weight of water saturated substrate mixture (g),  $W_d$ -weight of oven dried substrate mixture (g).

## Wettability $(mL \cdot L^{-1})$

Wettability was measured by soaking the pot filled with 1 L of oven dried substrate mixtures in standing water of 2 cm deep in plastic trays (Yahya et. al, 1997). The level of water was maintained by adding more water into the trays to ensure sufficient water for wetting the substrate mixtures. The degree of wettability was monitored hourly for 6 h by weighing the substrate mixture. The increase in moisture content with time was calculated by subtracting the wet weight of the substrate from its dry weight.

#### Air-filled porosity (%)

Air-filled porosity (AFP) was determined using saturation and drainage method at 2 h and 6 h after saturation. The pots filled with 1 L of the respective substrate mixtures were subirrigated by immersing them in the water. The substrate mixture was considered saturated when the water has appeared on the surface. The saturated substrate mixture was removed quickly to a funnel with a 500 mL cylinder. The volume of water drained from the pot was supposed to be replaced by an equivalent volume of air into the substrate mixture. The percentage of AFP calculated by the following formula:

AFP (Volume %) = 
$$\frac{Volume \text{ of water drained } (mL) \times 100}{Volume \text{ of substrate mixture } (mL)}$$

#### Vegetative growth and yield parameters

Main stem diameter at the 6th leaf point and approximately 12 cm from the ground level, stem length of four shoots for each plant, and leaf number per plant were measured at the end of the experiment. Fruit fresh weight, number of fruit per plant, blossom-end rot (%BER, by umber of fruit) and yield per plant were recorded during the experiment.

#### Leaves tissue analysis

At the end of the experiment, leaves were washed with deionized water and dried at 70 °C for 96 h for determination of mineral compositions. The dried leaf samples were ground and digestion was done according to Rodushkin et al. (1999). NO<sub>3</sub>-N, PO<sub>4</sub><sup>3-</sup>, Ca<sup>2+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, SO<sub>4</sub><sup>2-</sup>, Na<sup>+</sup> and Cl<sup>-</sup> were analyzed using HPLC ion analyzer (IA 300, TOA DKK Corporation, Japan); for anion, PCI-205 column was used and mobile phase of 1.8 mM Na<sub>2</sub>CO<sub>3</sub> and 1.7 mMNaHCO<sub>3</sub> at 1.0 mL·min<sup>-1</sup> were used; for cation, PCI-322 column was used and mobile phase of 6 mM Methanesulfonic acid at 1.0 mL·min<sup>-1</sup> were used. Manganese (Mn) and iron (Fe) were analyzed using RQflex® 10 (Merck chemicals, Germany). Zinc (Zn) was analyzed using spectrophotometer (DR 2800, HACH Company, USA) at 620 nm.

#### Statistical analysis:

Data were analyzed separately by one-way analysis of variance (ANOVA) for growing substrate mixture properties and two-way ANOVA for growth and yield parameters using SPSS version 16.0 for Windows and the differences among means were determined by Tukey's test at  $P \le 0.05$ .

### **3. Results and Discussion**

#### **Properties of substrate mixtures**

#### Initial pH and EC

 Table 1. Initial pH and EC of four substrate mixtures (before planting).

Substrate mixtures	pH	EC ( $dS \cdot m^{-1}$ )
M <sub>1</sub>	6.34 c <sup>z</sup>	0.08 b
$M_2$	6.13 c	0.10 b
M <sub>3</sub>	7.38 a	0.19 a
M4	6.68 b	0.11 b
Р	< 0.001	< 0.001

<sup>2</sup>Means with different letter is significantly different by Tukey's test at  $P \le 0.05$ .  $M_1 = 50\%$  coco peat + 45% bora + 5% shodo,  $M_2 = 50\%$  coco peat + 45% perlite + 5% shodo,  $M_3 = 50\%$  carbonized rice husk + 45% perlite + 5% shodo,  $M_4 = 50\%$  sawdust + 45% perlite + 5% shodo. P represents the level of significance of one-way ANOVA.

The initial pH and EC are two important properties of any growing substrate mixture as these parameters directly influence the availability and indicate inherent nutrients status in the substrates, respectively. Variations in the mixture components markedly affected the initial pH and EC among the substrate mixtures (Table 1). The highest pH was recorded in M<sub>3</sub> treatment followed by others. Meanwhile, the lowest pH was recorded in M<sub>2</sub>, which was statistically similar to that of M<sub>1</sub>. Blom (1983) stated that most of the plants grow best in slightly acidic pH ranges of 6.2- 6.8 in soil based substrate formulations and 5.4-6.0 in soilless substrate. Furthermore, different plant species (and cultivars) have different pH range for optimal growth but overall the optimum pH of the soilless substrate for adequate availability of essential plant nutrients is around 6.0 (Yahya et al., 2009). Our result revealed that the initial pH of the substrate mixtures of M<sub>3</sub> and M<sub>4</sub> were relatively higher than optimum and that could be optimized by mixing of acid based fertilizers before planting. Meanwhile, M1 and  $M_2$  had optimum pH level for growth of sweet pepper. The carbonized rice husk ( $M_3$  treatment) is alkaline in nature owing to its higher potassium content. Very low pH values could result in toxic concentration of ions such as aluminum, zinc and copper, while chemical bindings can occur at pH above 7.5 (Nappi and Barberis, 1993). All these phenomena lead to nutrients unavailability to the plants. The optimum pH of container substrate formulations differs with plant species, but a pH of 5.0-6.5 can be tolerated by most of the plants (DeBoodt and

Verndonck, 1972; Hans et al., 2005). Our present result agreed with these results.

Regarding EC, M<sub>3</sub> possessed the highest initial EC (0.191 dS m<sup>-1</sup>), while M<sub>1</sub> had the lowest EC (0.082 dS m<sup>-1</sup>), which was statistically similar to that of others. The EC values reflected the total inorganic ion concentration in the extracts of substrate mixtures. Higher EC of M<sub>3</sub> reflected that carbonized rice husk contained relatively higher concentration of soluble salts, which could support initial plant growth. Similarity to our results, Yahya et al. (2009) reported the higher initial EC of burnt rice hull mixture. High EC above 3.5 mS cm<sup>-1</sup> in substrate formulations causing poor plant growth (Hans et al., 2005 ; Lemaire et al., 1985; Eames, 1977). EC value below 2.0 mS cm<sup>-1</sup> is generally considered optimal to support the plant growth in container production systems (Milks et al., 1989). In our experiment, EC values for all the treatments did not exceed the higher values of EC. The results also indicated that EC value of M<sub>3</sub> treatment could help better initial growth of sweet pepper, but the initial pH value for this substrate was bit higher than the optimum level and that could be minimized by applying acid based fertilizers or by incorporating weak acid before planting.

#### **Bulk** density

Table 2. Bulk density, water retention and dry weight of four substrate mixtures (before planting).

Substrate mixtures	Bulk density (g·cm <sup>-3</sup> )	Water retention (%),	Dry weight $(g \cdot L^{-1})$
M <sub>1</sub>	0.41 a <sup>z</sup>	149.60 d	369.38 a
$M_2$	0.24 b	302.48 a	216.03 b
M <sub>3</sub>	0.20 c	269.78 b	202.38 b
M4	0.22 bc	248.48 c	195.18 b
Р	<0.001	< 0.001	< 0.001

<sup>z</sup>Means with different letters are significantly different by Tukey's test at  $P \le 0.05$ . *P* represents the level of significance of one-way ANOVA.

Bulk density differed significantly among the substrate mixtures (Table 2). The incorporation of bora and shodo into coco peat increased the bulk density ( $M_1$  treatment). Meanwhile the lowest bulk density was found in  $M_3$  treatment. Result obtained was consistent with the result by Islam (2008) who found that the bulk density of loose rice husk charcoal (burnt rice hull) was significantly lower than coco peat. The bulk density differed most likely due to the variation in particle-size distribution of the materials (Richards and Beardsell, 1986).

Bilderback et al. (2005) suggested that the acceptable range of bulk density for substrate is 0.19 to 0.70 g·cm<sup>-3</sup>. In our experiment, all the treatments had the bulk density within the acceptable range. Substrate mixture with low bulk density is required for frequently irrigated greenhouse to avoid oxygen deficiency. Furthermore, it can provide easier mixing and transportation than those of substrate with high bulk density. In this respect, substrate mixture containing carbonized rice husk (M<sub>3</sub>) could be more suitable for adequate aeration into the root zone of the plant.

#### Water retention

Results pertaining to water retention of four growing substrate mixtures are shown in Table 2. The volume of water held by different substrate mixtures differed significantly among the treatments. The highest water retention was recorded in  $M_2$  treatment and the lowest was in  $M_1$ . This might be due to its high proportion of macropores in which much of the water is lost by gravity. On the other hand,  $M_2$  containing high proportion of micropores had the highest water retention. Differences in available water holding capacity of the substrate could be due to their total porosity and type of pores (Bunt, 1988). Loss of water through gravity forces can be reduced incorporating finer particle into the substrates. Sambo et al. (2008) reported that substrate composed of ground rice hull with smaller particle size had smaller total pore space, and it contained more available water. But in our experiment,  $M_2$  treatment contained finer particles than other treatments. Furthermore, the second highest water holding capacity of  $M_2$  and  $M_3$  were comparatively greater and it can help in sustaining root development by releasing nutrients to the plant as and when needed.

#### Dry weight of substrate mixtures

Significant variation was found among different treatments for dry weight of substrate mixtures (Table 2). The highest dry weight observed in M<sub>1</sub> treatment, while the lowest in M<sub>4</sub>. Dry weight of substrate is an important criterion for easy mixing and transportation. It also affects construction materials for soilless culture. The grower can make hydroponic structure with low-cost materials, if the dry weights of substrate mixtures become low. The results obtained from the present study indicated that carbonized rice husk (M<sub>3</sub>) and sawdust (M<sub>4</sub>) based substrate can facilitate the growers to construct hydroponic structure with low-cost materials. Furthermore, it can help easy mixing and reusing as growing substrate.

## Air-filled porosity

Substrate mixtures	Air-filled porosity (%) at different times after drainage						
Substrate minitales	2 h	5 h					
M <sub>1</sub>	26.88 a	28.85 a					
M <sub>2</sub>	21.80 b	23.33 b					
<b>M</b> <sub>3</sub>	16.69 c	17.71 c					
<b>M</b> 4	23.67 b <sup>z</sup>	25.30 b					
Р	<0.001	<0.001					

Table 3. Air-filled porosity (AFP) of four substrate mixtures at 2 and 5 h after drainage.

<sup>z</sup>Means with different letter is significantly different by Tukey's test at  $P \le 0.05$ . P represents the level of significance of one-way ANOVA.

The air-filled porosity (%AFP) of the substrate mixtures differed significantly at both of 2 h and 5 h after drainage (Table 3). The substrate mixture of M<sub>1</sub> had significantly higher %AFP than that of other substrate mixtures at both of 2 h and 5 h after drainage. Meanwhile, the lowest %AFP was found from M<sub>3</sub> treatment. This result indicated that water rapidly removed by gravitational forces from M<sub>1</sub> treatment after saturated condition. Aeration depends mainly on the size of pores in a substrate mixture (Yahya et al., 2009). Water remains in the small pores after drainage by gravitational force, while large pore becomes empty and it is filled with air. Increasing the proportion of large pores allows more aeration after drainage has stopped (Handreck and Black, 2007). Richards and Beardsell (1986) found that exclusion of particles greater than 2 mm from a mixture of pine bark: sand: brown coal improved total water, available water and days to wilting without creating unfavorable level of aeration. Our result agreed with these results and it was indicated that M<sub>3</sub> treatment had higher proportion of small pore spaces that improved available water for the plant and decreased %AFP.

#### Wettability

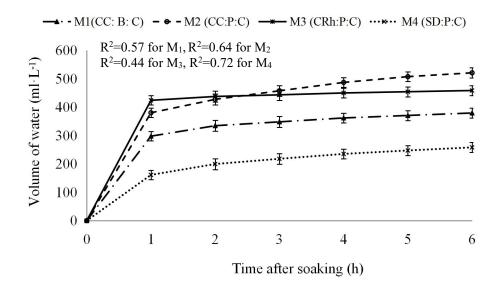


Fig. 1. Wettability of four substrate mixtures at different times after soaking. Vertical bars represent the standard error of the treatment means.

The wettability of the substrate mixture differed significantly and behaved differently over duration of soaking (Fig. 1). The highest water absorbing capacity after 1 h of soaking was observed in M<sub>3</sub>, which was followed by M<sub>2</sub> with the value of 423.75 mL·L<sup>-1</sup> and 379.92  $mL \cdot L^{-1}$ , respectively. The wettability of all the treatments increased over duration of soaking and the highest wettability was found in M<sub>2</sub>, which was followed by M<sub>3</sub> after 6 h of soaking. The higher amount of water absorbed by these two substrate mixtures ( $M_2$  and  $M_3$ ) reflected the synergistic effects of carbonized rice husk and coco peat when they were incorporated into perlite and shodo. Meanwhile, mixture of sawdust and coco peat with perlite and bora respectively had the lowest moisture content at 6 h after soaking suggesting that M<sub>1</sub> and M<sub>4</sub> contained high proportion of macropores. Considering the results observed on wettability, M2 and M<sub>3</sub> could be wetted easily and could supply water rapidly after irrigation to the plants grown on them. Yahya et al. (2009) observed the highest wettability in 70% coco: 30% burned rice husk followed by 70% coco: 30% perlite. Our result agreed with this result. Wettability is particularly important property for growing substrate, since it determines initial water uptake of the substrate and their subsequent water movement following root water removal and evapotranspiration (Michel et al., 2001). Therefore, the result indicated that M2 and M3 are considered as good growing substrate for horticultural crop production.

#### Plant growth and yield components

#### Vegetative growth

pepper.								
Trea tme nt	Main stem diamete r (mm)	Stem length (cm)	Number of leaf per plant	Leaf area per plant (cm <sup>2</sup> )	Fruit fresh weight (g)	Number of fruit per plant	BER (%) by number	Yield per plant (g)
Subst	trate mixtu	re (M)						
$M_1$	17.13 c <sup>z</sup>	661.83 c	129.00 b	2086.33 b	191.77 c	17.31 b	12.61 b	2242.1 b
$M_2$	18.08 b	707.50 b	135.00 a	2236.66 a	203.96 b	22.15 a	11.50 b	2619.5 a
M <sub>3</sub>	20.26 a	774.83 a	133.00 a	2286.67 a	210.84 a	21.07 a	9.48 c	2666.2 a
M4	15.17 d	621.00 d	116.83 c	1885.33 c	188.47 c	15.22 c	17.50 a	1961.5 c
Culti	vars (V)							
$\mathbf{V}_1$	18.92 a	667.75 b	135.75 a	2232.92 a	148.44 b	27.9 a	12.25 b	2681.8 a
$V_2$	16.40 b	714.83 a	121.17 b	2014.58 b	249.09 a	9.98 b	13.29 a	2062.8 b
Р								
М	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001
V	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	0.002	< 0.001
M× V	0.020	0.001	<0.001	0.005	NS	0.004	NS	0.005

 Table 4. Effect of different substrate mixtures and cultivars on vegetative growth and yield parameters of sweet

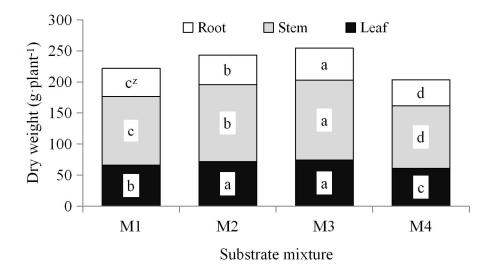
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<sup>2</sup>Means with different letter is significantly different by Tukey's test at  $P \le 0.05$ . *P* represents the level of significance of ANOVA. V<sub>1</sub> = Papri new-E-red and V<sub>2</sub> = AVRDC PP046-6006.

Vegetative growth of sweet pepper was significantly affected by different growing substrate mixtures, cultivars and their interactions (Table 4). Results showed that the vegetative growth tended to increase in sweet pepper grown in M<sub>3</sub> treatment. The highest main stem diameter, stem length, and leaf area were found from M<sub>3</sub>. On the other hand, the highest number of leaf was found in M<sub>2</sub> treatment, which was similar to that of M<sub>3</sub> treatment. These results revealed that M<sub>2</sub> and M<sub>3</sub> treatments had better properties as growing substrate for sweet pepper production. This might be due to proper aeration, water holding capacity, lower bulk density

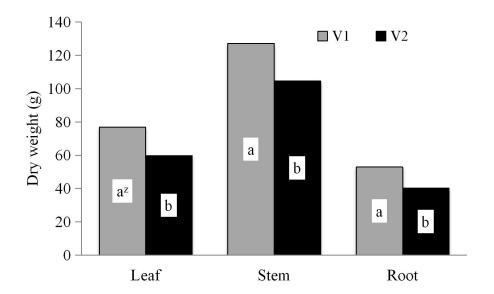
and biostability of carbonized rice husk (M<sub>3</sub> treatment) as compared to other treatments. Lemaire (1995) reported that lack of biostability may cause severe volume loss resulting in compaction, reduction in air volume, readily available water, and porosity due to mineralization and also changes in gaseous phase composition due to carbon dioxide production. These changes may finally reduce the plant growth (Lemaire, 1995). Thus, the lower plant growth was observed in sawdust (M<sub>4</sub> treatment) due to these changes in properties of mixture. High content of lignin in coconut coir was reported by Abad et al. (2002) and thus the coconut coir is most likely to be more biostable. On the other hand, carbonized rice husk is more sterile than the other substrate mixture and it also contains more nutrients. Therefore, better vegetative growth of sweet pepper was found in M<sub>3</sub> treatment. Yahya et al (2009) found the highest plant growth of *Celosia cristata* grown in burnt rice husk based growing substrate. Our results agreed with these findings.

On the other hand, the highest main stem diameter, number of leaves and leaf areas was found in  $V_1$  but stem length was higher in  $V_2$ . The results showed that  $V_1$  performed better in respect to stem diameter, number of leaves and leaf area. Jovicich et al. (2004) found the taller plants at densely planted sweet pepper, but density in our experiment was not higher than the commercially grown pepper.



#### Plant dry weight

Fig. 2. Effect of different substrate mixtures on plant dry weight of sweet pepper. <sup>z</sup>bars with different letter is significantly different among the vertical bars by Tukey's test at P≤0.05.



Different parts of plant

## Fig. 3. Effect of cultivars on plant dry weight of sweet pepper. <sup>z</sup>bars with different letter is significantly different between the vertical bars by Tukey's test at P≤0.05.

Dry weight in different parts of sweet pepper plant was significantly varied by substrate mixtures, cultivars and their interaction (Fig. 2 and 3). Significantly lower dry weight in different parts of the plants was found from M<sub>4</sub> treatment followed by others. Meanwhile the highest dry weight was found from M<sub>3</sub>. Overall, plants grown on M<sub>3</sub> were the heaviest and this is mainly associated with their leaf, stem and root dry weights. Results indicated that dry weight of plants grown on M<sub>2</sub> and M<sub>3</sub> were similar. Better root growth of plants grown on M<sub>2</sub> and M<sub>3</sub> could be attributed to the greater water availability and favorable aeration following the incorporation of burnt rice hull and coco peat into perlite. Under such condition, plants were provided with a sufficient water and oxygen. Furthermore, water availability and aeration, better root initiation and development could also be due to darker environment provided by burnt rice hull (Yahya et al. 2009). Darker rhizosphere environment was reported to promote translocation and accumulation of auxin at the basal part of the plant and thus a faster cell division and differentiation for root formation (Bilderback and Lorscheider, 1997). Similar to our results, Yahya et al. (2009) reported the heaviest plant dry weight of *Celosia cristata* grown in burnt rice hull.

Our result revealed that  $V_1$  produced higher dry weight compared to  $V_2$ . It might be due to higher vegetative growth in  $V_1$ .

#### Yield and yield components

Fruit fresh weight, number of fruit per plant, %BER and yield of sweet pepper were significantly varied by substrate mixture, cultivars and their interaction (Table 4). The plants in M<sub>3</sub> produced the highest fruit fresh weight and yield per plant. Meanwhile, the plants in M<sub>2</sub> produced the highest number of fruit which was statistically not different from that of M<sub>3</sub>. On the contrary, M<sub>4</sub> produced the lowest yield and yield parameters. It revealed that M<sub>2</sub> or M<sub>3</sub> substrate mixture was better than other treatments. This might be due to their improved physical and chemical properties discussed. Furthermore, the lowest incidence of %BER was found in M<sub>3</sub>.

Regarding cultivars, the highest number of fruit and yield, and lower incidence of %BER were found in  $V_1$ . Although  $V_2$  produced extra large, marketable fruit, but the total number of fruit was almost double in  $V_1$ . However,  $V_1$  performed better under different growing substrates. Rahman and Inden (2012) reported the highest yield and number of fruit in  $V_1$  compared to  $V_2$  under high temperature, which agreed with our present experiment using different growing substrates.

#### Mineral composition in leaves

Treatment		PO <sub>4</sub> <sup>3-</sup>	K	Ca	Mg	SO4 <sup>2-</sup>	Fe	Mn	Zn	
	g·kg <sup>-1</sup>							ppm		
Substrate r	Substrate mixture (M)									
$M_1$	6.75 b <sup>z</sup>	3.07 c	43.25 c	21.66 c	10.09 a	1.68 c	37.50	56.00	20.83	
$M_2$	7.35 a	3.36 b	47.12 b	23.12 b	9.16 c	1.85 b	38.00	54.83	21.00	
M <sub>3</sub>	7.65 a	3.57 a	51.01 a	24.36 a	9.67 b	2.03 a	37.33	54.33	20.17	
M4	5.50 c	2.75 d	39.01 d	19.81 d	9.55 b	1.52 d	37.17	54.50	20.16	
Cultivars (V)										
$V_1$	7.33 a	3.31 a	47.75 a	22.91 a	9.68 a	1.66 b	40.17 a	58.67 a	21.42 a	
$V_2$	6.30 b	3.06 b	42.74 b	21.56 b	9.59 b	1.88 a	34.83 b	51.17 b	19.67 b	
P										
М	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	ns	ns	ns	

V	< 0.001	< 0.001	< 0.001	< 0.001	0.003	< 0.001	< 0.001	< 0.001	< 0.001
$\mathbf{M}\times\mathbf{V}$	ns	ns	0.005	< 0.001	< 0.001	< 0.001	ns	ns	ns

<sup>z</sup>Means with different letter is significantly different by Tukey's test at  $P \le 0.05$ . *P* represents the level of significance of ANOVA. V<sub>1</sub> = Papri new-E-red and V<sub>2</sub> = AVRDC PP046-6006.

Macronutrients in leaves were significantly affected by substrate mixtures, cultivars and their interactions (Table 5). Meanwhile, micronutrient contents were insignificant by substrate mixtures and interactions, but significant for cultivars. Results indicated that all macro nutrient contents were the highest in M<sub>3</sub> and lowest in M<sub>4</sub>. This might be due to differences of physical and chemical properties among substrate mixtures. The mineral compositions in leaves recorded in this study were in the normal range found in healthy mature leaf tissue (Jobin, et al., 2004). The physical properties of substrate are important factor in determining plant development in soilless substrate (Karla, 1998; Michel et al., 2001; Chavez et al., 2008). Therefore, differences in the leaf macro nutrients, and variation in the growth and yield of sweet pepper observed in this study were linked with the differences in the chemical and physical properties of the substrate mixtures.

On the other hands, all nutrient compositions were the highest in  $V_1$  except for  $SO_4^{-2}$ . Variation in all nutrient composition in leaves improved the growth and yield of  $V_1$  cultivar as compared to  $V_2$  cultivar.

## 4. Conclusion

Results indicated that certain chemical and physical properties of carbonized rice husk (M<sub>3</sub>) and coco peat (M<sub>2</sub>) can be improved through incorporation of perlite and shodo. The positive effects of carbonized rice husk based substrate mixture (M<sub>3</sub>) were found on the elevation of nutrient availability (as indicated by higher EC), bulk density; lower air-filled porosity, high available water and wettability. Improvement in chemical and physical properties of carbonized rice husk (M<sub>3</sub>) and coco peat (M<sub>2</sub>) based growing substrate mixtures resulted in better plant growth and yield of sweet pepper. On the contrarily, improved physical and chemical properties were not found in saw dust (M<sub>4</sub>) based substrate mixture that negatively affected the growth and yield of sweet pepper. Therefore, it is suggested that carbonized rice husk (M<sub>3</sub>) can be used for better growth and yield of sweet pepper 'Papri new-E-red' cultivar.

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